

Genetic and environmental determinants for disease risk in subsets of rheumatoid arthritis defined by the anticitrullinated protein/peptide antibody fine specificity profile

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ABSTRACT

Objectives To increase understanding of the aetiology and pathogenesis of rheumatoid arthritis (RA), genetic and environmental risk factors for RA subsets, defined by the presence or absence of different anticitrullinated protein/peptide antibodies (ACPAs) targeting citrullinated peptides from α -enolase, vimentin, fibrinogen and collagen type II, were investigated.

Methods 1985 patients with RA and 2252 matched controls from the EIRA case-control cohort were used in the study. Serum samples were assayed by ELISA for the presence of anticyclic citrullinated peptides (anti-CCP) antibodies and four different ACPA fine specificities. Cross-reactivity between ACPAs was examined by peptide absorption experiments. Genotyping was performed for *HLA-DRB1* shared epitope (SE) alleles and the *PTPN22* gene, while information regarding smoking was obtained by questionnaire. The association of genetic and environmental risk factors with different subsets of RA was calculated by logistic regression analysis.

Results Limited cross-reactivity was observed between different ACPA fine specificities. In total, 17 RA subsets could be identified based on their different ACPA fine specificity profiles. Large differences in association with genetic and environmental determinants were observed between subsets. The strongest association of *HLA-DRB1* SE, *PTPN22* and smoking was identified for the RA subset which was defined by the presence of antibodies to citrullinated α -enolase and vimentin.

Conclusion This study provides the most comprehensive picture to date of how *HLA-DRB1* SE, *PTPN22* and smoking are associated with the presence of specific ACPA reactivities rather than anti-CCP levels. The new data will form a basis for molecular studies aimed at understanding disease development in serologically distinct subsets of RA.

INTRODUCTION

Rheumatoid arthritis (RA) has for many years been considered a prototype for immune-mediated diseases owing to the strong genetic association with certain MHC class II alleles (ie, *HLA-DRB1* shared epitope (SE)).^{1–3} More recently the MHC class II association in RA was shown to be confined to a subset of patients characterised by the presence of antibodies to synthetic cyclic citrullinated peptides

(CCP).^{4,5} The concomitant recognition that the best known environmental risk factor for RA (ie, smoking) constitutes a risk only for the development of CCP-positive disease, together with the finding that smoking induces the exposure of citrullinated proteins in the lungs,^{4,6,7} suggests an aetiology for RA where *HLA-DRB1* SE-restricted immune reactions to citrullinated epitopes trigger and drive disease pathogenesis.⁸ Subsequently, it has been shown that the CCP assays such as the most frequently used CCP2 ELISA is able to capture anticitrullinated protein/peptide antibodies (ACPAs) with a broad spectrum of fine specificities including naturally occurring citrullinated autoantigens.^{9,10}

In order to increase our understanding of the aetiology and pathogenesis of RA it has therefore become important to define which ACPA fine specificities are associated with the different *HLA-DRB1* SE alleles. Such studies have been performed for some ACPA fine specificities^{11–15} and differences within the CCP-positive population have been described. However, no comprehensive picture has emerged to date, partly because only single target antigens have been studied and partly due to small study populations and lack of information on appropriate controls. In the present study we have used the large and well-characterised Epidemiological Investigation of Rheumatoid Arthritis (EIRA) case-control cohort¹⁶ to investigate *HLA-DRB1* SE in combination with *PTPN22* polymorphism—the second most important genetic risk factor for CCP-positive RA¹⁷—and cigarette smoking in relation to the presence of antibodies targeting four well-defined citrullinated autoantigens (citrullinated α -enolase, -vimentin, -fibrinogen and -collagen type II).

METHODS

Subjects

EIRA is an ongoing Swedish population-based case-control study which is described in detail elsewhere.¹⁶ In the present study we analysed 1985 incident cases of RA included in the EIRA study between May 1996 and May 2006 and 2252 controls randomly selected from the national population register to match RA cases in terms of age, gender and residential area. Cases were diagnosed with RA according to the 1987 American College



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of Rheumatology criteria.¹⁸ Cases and controls donated blood at the time of inclusion and completed a self-administered questionnaire relating to life style and environmental exposures.

ELISA

Anti-CCP antibody status was determined in serum from 1985 cases and 1210 controls using the CCP2 assay (Immunoscan CCPlus, Euro-Diagnostica, Malmö, Sweden) according to the manufacturer's instructions. The cut-off for positivity was 25 AU/ml. Antibody responses to citrullinated peptides from α -enolase (CEP-1), vimentin (Cit-vim), fibrinogen (Cit-fib) and collagen type II (Cit-C1) or the arginine-containing equivalents were assayed in serum samples from 1985 EIRA cases and 150 randomly selected EIRA controls using in-house ELISAs, as previously described.^{11 19} The cut-off for positivity was 10 AU/ml for all ACPA fine specificities. A detailed description of the ELISA method is given in the online supplement.

Cross-reactivity assay

Ten serum samples with high anti-CCP levels (>800 AU/ml) that also displayed reactivity to CEP-1 (n=10), Cit-vim (n=8), Cit-fib (n=9) and Cit-C1 (n=8) were selected from EIRA for cross-reactivity experiments. Each serum (diluted 1:100) was incubated with each of the four peptides (CEP-1, Cit-vim, Cit-fib or Cit-C1) at 10 μ g/ml or in the presence of buffer alone. Preabsorption was performed in liquid phase during constant agitation for 2 h at room temperature. Following centrifugation (15 min, 1000 g), the supernatants were transferred to the four peptide ELISA plates and assayed as described above. Inhibition was determined based on the differences in AU/ml values between the absorbing peptide and buffer alone.

Peptides

Freeze-dried synthetic peptides (>95% purity) corresponding to sequences from human α -enolase (amino acid 5–21), vimentin (amino acid 60–75) and fibrinogen (amino acid 36–52) in its 'native' form or with arginine to citrulline substitutions (Innovagen, Lund, Sweden), were dissolved at 10 mg/ml (Milli-Q water, 10% DMSO) and stored at -20°C before used. The homotrimeric triple helical collagen type II peptides C1 and Cit-C1 were synthesised as previously described.^{20 21} A detailed description of the peptide sequences is given in table 1 in the online supplement.

Genotyping

HLA-DRB1 subtyping was performed on DNA samples from 1961 RA cases and 1278 controls by sequence-specific primer PCR, as previously described.¹¹ *DRB1*01* (except *DRB1*0103*), *DRB1*04* and *DRB1*10* were classified as shared epitope (SE) alleles of *HLA-DRB1*. The protein tyrosine phosphatase gene (*PTPN22* rs2476601) was genotyped on 1943 RA cases and 1284 controls, as previously described.¹⁷

Cigarette smoking

Information regarding cigarette smoking prior to the appearance of symptoms of arthritis in 1727 RA cases and the corresponding time points among 1913 controls were obtained by questionnaire. Individuals were categorised as ever smokers (current/past) or never smokers.¹⁶

Statistical analyses

To determine the association of *HLA-DRB1* SE (alone or in combination with *PTPN22* and/or smoking) with different subsets of RA, ORs with 95% CI were calculated using unconditional

logistic regression analyses, with cases and controls that were negative for the risk factors as the referent category. All analyses were performed with adjustment for design variables (age, residential area and gender).

Interaction, defined by departure from additivity of effects,²² was evaluated between *HLA-DRB1* SE and smoking and between *HLA-DRB1* SE and *PTPN22*. The attributable proportion due to interaction (AP) was calculated together with 95% CI, as previously described.^{11 23} The AP between two interacting factors reflects the joint effect beyond the sum of the independent effects. All analyses were performed using SAS Version 9.1.

Differences in ACPA levels between different RA subsets or between absorbing peptides and buffer in the cross-reactivity experiment were examined using the Mann–Whitney U test for independent groups.

RESULTS

Identification of different RA subsets based on ACPA fine specificity profiles

Sixty-three percent of RA cases in EIRA were anti-CCP antibody positive (35% anti-CEP-1 positive, 37% anti-Cit-vim positive, 28% anti-Cit-fib positive and 37% anti-Cit-C1 positive). Low-level antibody responses to the arginine-containing control peptides from α -enolase, vimentin and fibrinogen were detected in less than 2.4% of patients, while reactivity to the arginine-containing C1 peptide^{20 21 24} of collagen type II was detected in 9% (data not shown). The presence of different ACPA fine specificities overlapped to a large extent and, by analysing the different patterns, 17 serologically distinct subsets could be

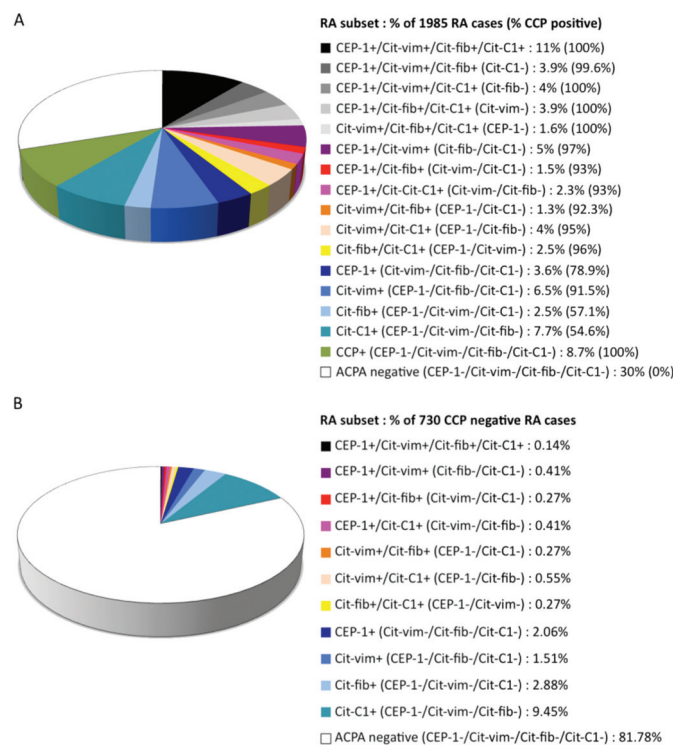


Figure 1 Subsets of rheumatoid arthritis (RA) defined based on the presence (+) or absence (-) of four different anticitrullinated protein/peptide antibody (ACPA) fine specificities (α -enolase (CEP-1), vimentin (Cit-vim), fibrinogen (Cit-fib) and collagen type II (Cit-C1)) as detected by ELISA in (A) 1985 cases with RA and (B) 730 anticyclic citrullinated peptides (anti-CCP) negative RA cases. In (A) the subsets may comprise CCP positive and CCP negative RA cases, with the percentages of anti-CCP positive cases shown in brackets.

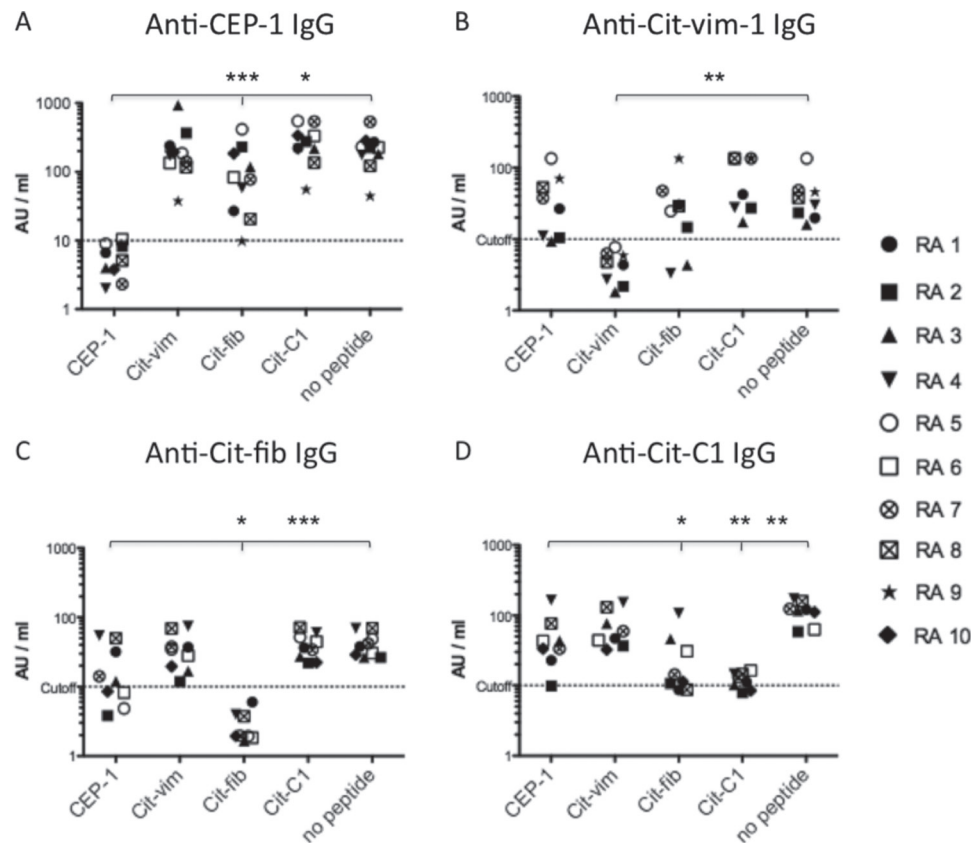


Figure 2 Cross-reactivity between different anticitrullinated protein/peptide antibody (ACPA) fine specificities. Antibody responses to citrullinated peptides from α -enolase (CEP-1), vimentin (Cit-vim), fibrinogen (Cit-fib) and collagen type II (Cit-C1) were measured in serum samples from 10 patients with cyclic citrullinated peptide (CCP) positive rheumatoid arthritis (RA1–RA10) after preabsorption with the different peptides: (A) anti-CEP-1 IgG levels; (B) anti-Cit-vim IgG levels; (C) anti-Cit-fib IgG levels; (D) anti-Cit-C1 IgG levels. Serum samples that were negative for one or several ACPA fine specificities were removed from those specific analyses: RA6 and RA10 (B); RA9 (C); RA5 and RA9 (D). Log scales of AU/ml values are shown on the y-axis. Dotted lines indicate cut-off for positivity (10 AU/ml). The x-axis indicates preabsorbing peptides or no peptide (buffer alone). *** $p < 0.0001$, ** $p < 0.001$, * $p < 0.05$ vs no peptide.

identified (figure 1A). Most of the patients who were positive for the specific ACPAs were confined to the CCP-positive population, although 18% of CCP-negative patients were positive for at least one ACPA fine specificity (figure 1B). Conversely, 8.7% of all patients were CCP-positive but negative for the four ACPA fine specificities.

In the CCP-negative population, ACPA-positive patients typically showed reactivity to only one citrullinated antigen where anti-Cit-C1 antibodies were dominant (9.45%). In the CCP-positive subset, on the other hand, the presence of two (16.6%), three (13.4%) or four (11%) ACPA fine specificities was common. ACPA fine specificity levels were generally lower in the CCP-negative population than in the CCP-positive population (data not shown). Moreover, there was a strong correlation between the number of ACPAs present and anti-CCP antibody levels, with a significant ($p < 0.0001$) increase in anti-CCP antibody levels for every additional ACPA fine specificity (see figure 1 in online supplement).

ACPA show limited cross-reactivity

In order to analyse the extent of cross-reactivity between antibodies reactive with the different citrullinated peptides, serum samples from 10 CCP-positive RA patients with antibodies to CEP-1 ($n=10$), Cit-vim ($n=8$), Cit-fib ($n=9$) and Cit-C1 ($n=8$) were preabsorbed against each of the four peptides before being retested for ACPA reactivities. All four ACPAs were efficiently

absorbed by preincubation with its homologous peptide, while only limited cross-reactivity was observed between specificities. Anti-CEP-1 IgG showed a low degree of cross-reactivity with Cit-fib in some serum samples ($p=0.043$) but no cross-reactivity with Cit-vim or Cit-C1 (figure 2A). Anti-Cit-vim IgG showed cross-reactivity with Cit-fib in two serum samples (figure 2B). Anti-Cit-fib IgG showed some degree of cross-reactivity with CEP-1 in some serum samples ($p=0.05$, figure 2C) and anti-Cit-C1 antibodies showed cross-reactivity mainly with Cit-fib in approximately half of the serum samples ($p < 0.001$, figure 2D).

HLA-DRB1 SE in combination with PTPN22 and smoking is associated mainly with CEP-1 and Cit-vim positive RA

To investigate associations of genetic and environmental risk factors with different subsets of RA, we first defined RA subsets by the presence or absence of antibodies to single citrullinated peptides—that is, without taking into account which additional ACPA fine specificities may also be present in the subset. As shown in figure 3A, being positive for *HLA-DRB1* SE, *PTPN22* and smoking was associated mainly with CEP-1 positive RA (OR 39.1) compared with CEP-1 negative/CCP positive RA (OR 7.9). A similar pattern was observed for antibodies to Cit-vim where the association of *HLA-DRB1* SE in combination with *PTPN22* and smoking was stronger with Cit-vim positive disease OR 25.6 compared with Cit-vim negative/CCP positive

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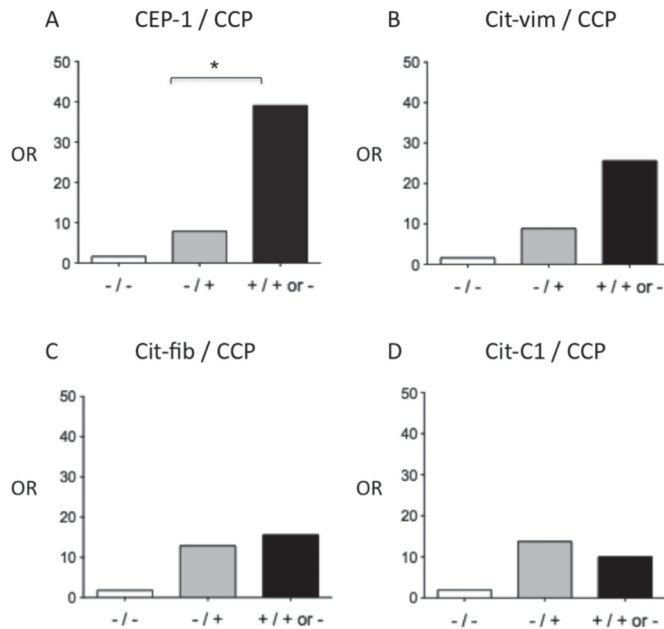


Figure 3 Combined effect of *HLA-DRB1* SE, *PTPN22* and smoking on disease risk in different subsets of rheumatoid arthritis (RA). ORs with 95% CI were calculated using unconditional logistic regression analysis by comparing individuals positive for the risk factors with individuals who were negative. RA subsets were defined based on the presence or absence of one anticitrullinated protein/peptide antibody (ACPA) fine specificity without taking into account which other ACPA fine specificities may be present simultaneously. Antibody responses to citrullinated peptides from (A) α -enolase (CEP-1), (B) vimentin (Cit-vim), (C) fibrinogen (Cit-fib) and (D) collagen type II (Cit-C1). White bars show subsets negative for the ACPA fine specificity and negative for cyclic citrullinated peptide (CCP) (-/-); grey bars show subsets negative for the ACPA fine specificity but positive for CCP (-/+); black bars show subsets positive for the ACPA fine specificity but where the CCP status was not considered (+/+ or -). 95% CIs for -/+ and +/+ or - subsets are: (A) 4.58 to 13.53 vs 19.07 to 79.97; (B) 5.03 to 15.68 vs 13.95 to 47.02; (C) 7.55 to 0 vs 8.51 to 4; (D) 7.55 to 9 vs 6.07 to 9. Asterisks indicate non-overlapping 95% CI.

Table 1 ORs for disease risk in different subsets of rheumatoid arthritis in carriers of *HLA-DRB1* shared epitope (SE) compared with non-carriers

ACPA	CEP-1/Cit-vim/Cit-fib/Cit-C1	<i>HLA-DRB1</i> SE		OR*	95% CI
		Any	None		
Control group	NA	643	635	1.0	Ref
Case groups	+/-/-/-	59	12	5.1	2.7 to 9.5
	-/+/-/-	108	18	6.0	3.6 to 10.0
	-/-/+/-	28	21	1.4	0.75 to 2.5
	-/-/-/+	91	57	1.6	1.11 to 2.3
	+/-/+/-	97	2	49.6	12.1 to 202.5
	+/-/+/-	21	7	3.2	1.3 to 7.6
	+/-/-/+	43	3	15.0	4.6 to 49.0
	-/+/+/-	18	8	2.3	1.0 to 5.4
	-/+/-/+	69	10	6.9	3.5 to 13.6
	-/-/+/+	36	14	2.5	1.3 to 4.7

*Adjusted for age, gender and residence area.
ACPA, anticitrullinated protein/peptide antibodies.

disease OR 8.9, figure 3B). Subsets defined by antibodies to Cit-fib or Cit-C1, on the other hand, did not demonstrate such specific associations beyond the effect of CCP (OR 15.6 vs 12.9 for Cit-fib and 10.0 vs 13.8 for Cit-C1; figure 3C,D).

Table 2 ORs for disease risk in different subsets of rheumatoid arthritis in subjects exposed to different combinations of *HLA-DRB1* shared epitope (SE) and smoking compared with non-exposed subjects

CEP-1/Cit-vim/ Cit-C1	<i>HLA-DRB1</i> SE	Smoking	Cases/controls	OR*	95% CI
-/-/-	-	-	115/228	1	Ref
-/-/-	+	-	176/244	1.4	1.1 to 1.9
-/-/-	-	+	173/316	1.1	0.8 to 1.5
-/-/-	+	+	233/315	1.6	1.2 to 2.1
+/-/-	-	-	3/228	1	Ref
+/-/-	+	-	25/244	8	2.4 to 27.0
+/-/-	-	+	13/316	3.3	0.9 to 11.9
+/-/-	+	+	46/315	12.5	3.8 to 40.9
-/+/-	-	-	7/228	1	Ref
-/+/-	+	-	42/244	5.8	2.5 to 13.3
-/+/-	-	+	15/316	1.7	0.7 to 4.3
-/+/-	+	+	64/315	7.3	3.2 to 16.4
-/-/+	-	-	35/228	1	Ref
-/-/+	+	-	41/244	1.1	0.7 to 1.8
-/-/+	-	+	31/316	0.6	0.4 to 1.1
-/-/+	+	+	68/315	1.4	0.9 to 2.3
+/+/-	-	-	5/228	1	Ref
+/+/-	+	-	29/244	5.5	2.1 to 14.5
+/+/-	-	+	7/316	1	0.3 to 3.3
+/+/-	+	+	118/315	18.4	7.3 to 46.1
+/-/+	-	-	3/228	1	Ref
+/-/+	+	-	25/244	7.7	2.3 to 26.1
+/-/+	-	+	11/316	2.7	0.7 to 10.0
+/-/+	+	+	65/315	17.2	5.3 to 55.8
-/+/+	-	-	7/228	1	Ref
-/+/+	+	-	27/244	3.5	1.5 to 8.4
-/+/+	-	+	10/316	1	0.4 to 2.8
-/+/+	+	+	52/315	5.6	2.5 to 12.7
+/+/+	-	-	6/228	1	Ref
+/+/+	+	-	54/244	8.5	3.6 to 20.1
+/+/+	-	+	14/316	1.6	0.6 to 4.3
+/+/+	+	+	189/315	23.3	10.1 to 53.7

*Adjusted for age, gender and residential area.

Impact of *HLA-DRB1* SE on disease risk in different RA subsets

The results described above may be influenced by the coexistence of multiple ACPA fine specificities. We next investigated the four RA subsets which were defined by the sole presence of one ACPA fine specificity—that is, the subsets that were ‘single positive’ for anti-CEP-1, anti-Cit-vim, anti-Cit-fib or anti-Cit-C1 antibodies. In this analysis we focused on *HLA-DRB1* SE, the risk factor with the strongest impact. We found a relatively strong association of *HLA-DRB1* SE with CEP-1 positive RA (OR 5.1) as well as with Cit-vim positive disease (OR 6.0) but not with Cit-fib positive disease (OR 1.4), and the association with Cit-C1 positive RA was weak (OR 1.6) (table 1). A significant association of *HLA-DRB1* SE (OR 2.5, 95% CI 1.8 to 3.6) could also be observed with the subset that was positive for CCP but negative for all four ACPA fine specificities (data not shown).

We then studied the effect of being positive for different combinations of two ACPA fine specificities as the impact of *HLA-DRB1* SE may differ between subsets defined by specific combinations of non-cross-reactive ACPAs. The strongest influence of *HLA-DRB1* SE was identified for CEP-1/Cit-vim double-positive RA (OR 49.6) which could be compared with an OR of 2.5 for the association with Cit-fib/Cit-C1 double-positive RA. Interestingly, anti-CCP antibody levels were higher in the Cit-fib/Cit-C1 double-positive subset than in the CEP-1/Cit-vim double-positive subset, suggesting that *HLA-DRB1* SE mainly influence the fine specificity of the ACPA response rather than anti-CCP levels (see figure 2 in online supplement).

Table 3 ORs for disease risk in different subsets of rheumatoid arthritis in subjects exposed to different combinations of *HLA-DRB1* shared epitope (SE), smoking and *PTPN22* compared with non-exposed subjects

CEP-1 /Cit-vim	<i>HLA-DRB1</i> SE	Smoking	<i>PTPN22</i>	Cases/ controls	OR*	95% CI
-/-	-	-	-	106/178	1	Ref
-/-	-	-	+	41/49	1.4	0.9 to 2.3
-/-	+	-	-	168/200	1.4	1.0 to 1.9
-/-	+	-	+	47/43	2	1.2 to 3.2
-/-	-	+	-	140/233	1	0.8 to 1.4
-/-	-	+	+	61/81	1.4	0.9 to 2.1
-/-	+	+	-	223/252	1.6	1.1 to 2.1
-/-	+	+	+	76/61	2.4	1.5 to 3.6
+/-	-	-	-	4/178	1	Ref
+/-	-	-	+	2/49	1.9	0.3 to 10.7
+/-	+	-	-	32/200	7	2.4 to 20.3
+/-	+	-	+	18/43	20.1	6.4 to 63.2
+/-	-	+	-	13/233	2.6	0.8 to 8.2
+/-	-	+	+	10/81	5.7	1.7 to 18.8
+/-	+	+	-	80/252	15.4	5.5 to 43.1
+/-	+	+	+	29/61	24.4	8.2 to 73.1
-/+	-	-	-	10/178	1	Ref
-/+	-	-	+	4/49	1.7	0.5 to 5.7
-/+	+	-	-	47/200	4.3	2.1 to 8.8
-/+	+	-	+	22/43	9.9	4.3 to 22.8
-/+	-	+	-	21/233	1.7	0.8 to 3.8
-/+	-	+	+	4/81	1	0.3 to 3.3
-/+	+	+	-	83/252	6.3	3.2 to 12.7
-/+	+	+	+	33/61	11.5	5.3 to 25.3
+/+	-	-	-	6/178	1	Ref
+/+	-	-	+	5/49	3.4	1.0 to 11.7
+/+	+	-	-	43/200	6.4	2.7 to 15.6
+/+	+	-	+	40/43	30.4	12.0 to 76.9
+/+	-	+	-	14/233	1.8	0.7 to 4.8
+/+	-	+	+	7/81	2.7	0.9 to 8.2
+/+	+	+	-	212/252	26.6	11.5 to 61.7
+/+	+	+	+	92/61	50.1	20.6 to 121.8

*Adjusted for age, gender and residential area.

Impact of smoking in combination with *HLA-DRB1* SE on disease risk in RA subsets

Since the data presented above do not support an association of *HLA-DRB1* SE with the presence of anti-Cit-vim antibodies, we restricted the next analysis to RA subgroups defined by the presence or absence of antibodies to CEP-1, Cit-vim and Cit-C1. We had sufficient power to investigate the combined effect of *HLA-DRB1* SE and smoking. As shown in table 2, smoking in the absence of *HLA-DRB1* SE did not significantly associate with any RA subset but strengthened the *HLA-DRB1* SE association when present. This effect was most pronounced for double-positive and triple-positive subsets compared with single-positive subsets, but only when anti-CEP-1 antibodies were present. Adding Cit-vim reactivity to the CEP-1 single-positive subset increased the OR for the combined *HLA-DRB1* SE and smoking association from 12.5 to 18.4. A similar effect was seen when adding Cit-C1 reactivity to the CEP-1 single-positive subset, with an increase in the OR from 12.5 to 17.2. However, in the absence of anti-CEP-1 antibodies, addition of Cit-C1 reactivity to the Cit-vim single-positive subgroup did not increase the OR further (7.3 vs 5.6). The highest OR (23.3) for the combined *HLA-DRB1* SE and smoking association was observed for the triple-positive subset (CEP-1+/Cit-vim+/Cit-C1+).

Impact of *PTPN22* in combination with *HLA-DRB1* SE on disease risk in RA subsets

We then investigated the combined effect of *HLA-DRB1* SE and *PTPN22* in subsets defined by anti-CEP-1, anti-Cit-vim and anti-Cit-C1 antibody status. As seen with smoking, *PTPN22* also mainly had an effect in the presence of *HLA-DRB1* SE, and this association was also most pronounced for double-positive and triple-positive subsets encompassing anti-CEP-1 antibodies. Addition of Cit-vim reactivity to the CEP-1 single-positive subset increased the OR for the combined *HLA-DRB1* SE and *PTPN22* association from 8.7 to 29.2 while addition of Cit-C1 to the CEP-1 single positive subset changed the OR from 8.7 to 17.1. However, similar ORs were observed for the Cit-vim single-positive and the Cit-vim/Cit-C1 double-positive subsets (8.4 vs 8.7). The strongest association (OR 34.8) of the *HLA-DRB1* SE and *PTPN22* combination was seen with the triple-positive subset (CEP-1+/Cit-vim+/Cit-C1+) (see table 2 in online supplement).

Combined effect of *HLA-DRB1* SE, *PTPN22* and smoking on disease risk in RA subsets defined by anti-CEP-1 and anti-Cit-vim antibody status

In the final analysis we investigated the combined effect of all three risk factors (*HLA-DRB1* SE, *PTPN22* and smoking) and calculated gene-environment and gene-gene interactions. We focused on what appears to be the two most important ACPA fine specificities in this context and defined RA subgroups based on CEP-1 and Cit-vim reactivity alone (table 3). The presence of *HLA-DRB1* SE alleles, *PTPN22* polymorphism and cigarette smoking conferred a rather modest risk for the development of CEP-1 negative/Cit-vim negative disease (OR 2.4), while the risk was high for CEP-1 positive/Cit-vim negative RA (OR 24.4) and for CEP-1 negative/Cit-vim positive RA (OR 11.5). However, the highest risk was identified for the CEP-1/Cit-vim double-positive subset (OR 50.1).

Analyses of interaction, calculated as the AP,^{22 23} demonstrated significant interactions between *HLA-DRB1* SE and smoking in the CEP-1 positive/Cit-vim negative as well as in the CEP-1/Cit-vim double-positive subsets with AP values of 0.33 and 0.64, respectively. Interactions were also observed between *HLA-DRB1* SE and *PTPN22* in the CEP-1 positive/Cit-vim negative (AP=0.44), the CEP-1 negative/Cit-vim positive (AP=0.56) and in the CEP-1/Cit-vim double positive subsets (AP=0.53) (see table 3 in online supplement).

DISCUSSION

The present study provides novel data concerning the relationships between specific autoimmune reactions and genetic and environmental determinants for the risk of developing RA, with the overall conclusion that *HLA-DRB1* SE genes, the *PTPN22* risk allele and cigarette smoking are associated with the presence of specific ACPA-reactivities rather than total levels of anti-CCP antibodies.

We have focused our analyses on antibody responses to well-defined citrullinated epitopes on α -enolase,^{25 26} vimentin,^{13 27} fibrinogen^{28 29} and collagen type II,²¹ representing physiological antigens present in the rheumatoid joint. Most of the patients who were positive for these ACPA fine specificities were also positive in the CCP2 ELISA, the current standard assay for ACPA detection. Although important to note, 18% of the CCP negative patients were positive for at least one ACPA fine specificity. Conversely, nearly 14% of the CCP positive patients were negative for the four ACPA fine specificities, and this subset also

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showed some association with *HLA-DRB1* SE, clearly indicating that other ACPA targets exist, which has also been reported previously^{25 30 31} and should be considered in future studies.

Our peptide absorption experiments showed that the ACPAs were mainly non-cross-reactive. Although some degree of cross-reactivity was observed in some serum samples, the majority did not demonstrate cross-reactivity, in line with previous reports.^{9 14} We therefore consider these ACPAs as four distinct fine specificities, and we subsequently based our RA subsets on the presence or absence of anti-CEP-1, anti-Cit-vim, anti-Cit-fib and anti-Cit-C1 antibodies rather than CCP status. The fact that the ACPAs occur together in some patients, but on their own in others, imply that their production may be governed by partly different mechanisms. It is therefore very interesting to note the specific relationship between *HLA-DRB1* SE and certain ACPA fine specificities (particularly CEP-1 and Cit-vim) but not others (mainly Cit-fib).

The estimated risks in some of the analyses should be viewed in the context of small sample sizes with large CI. However, our data collectively point to clear differences between ACPA fine specificities, thus confirming and extending previous reports.^{11–13} Importantly, the present study answers the previously unanswered question concerning the influence of *HLA-DRB1* SE on the specificity versus the magnitude of the ACPA response.³² Also notable is the fact that the effects of smoking and *PTPN22*, in the context of *HLA-DRB1* SE, appear to be most pronounced in subsets defined by multiple ACPAs where reactivity to CEP-1 is present. These data may suggest that smoking is involved in citrullinating and exposing some—but not other—autoantigens to the immune system, and that *PTPN22* exerts its effects only when certain MHC class II-dependent immune reactions are involved. These data will form the basis for future experimental studies on molecular pathways of relevance for disease development in distinct subsets of RA.

In summary, our data highlight the complexity of the gene-environment influence on ACPA-positive RA. A particularly interesting finding is that *HLA-DRB1* SE together with *PTPN22* and smoking associate so strongly with the presence of anti-CEP-1 antibodies, on the one hand, and anti-Cit-vim antibodies on the other, but mainly with the simultaneous presence of both ACPAs. Since the SE comprises several different alleles and since a dose-effect has been demonstrated for the *HLA-DRB1* SE association,^{4 5 11} one may speculate that peptides from citrullinated α -enolase may be preferentially presented by one specific SE allele and subsequently trigger the production of anti-CEP-1 antibodies, while peptides from citrullinated vimentin are preferentially presented by another SE allele giving rise to anti-Cit-vim antibodies. Hence, in such a scenario, the presence of two different SE alleles would drive the simultaneous production of two different ACPAs which may tentatively synergise in contributing to the development of RA. Detailed studies of the SE alleles, other MHC class II alleles, as well as other genetic polymorphisms and environmental exposures in the context of different ACPA fine specificities will increase our understanding of the aetiology in different subsets of ACPA-positive RA. The results from such studies may also help to develop future personalised therapies for patients with RA and preventive strategies for individuals at risk of developing the disease.

Contributors KL had main responsibility for the study including design, experimental set-up, analyses, figures, supervising and writing the paper. CB had main responsibility for the statistical analyses and tables (association and interaction analyses). NK and ER performed ELISA assays, cross-reactivity experiments and produced figures with statistics. XJ produced the tables and, together with HK, contributed to the statistical analyses (association and interaction analyses). IP-D

performed ELISA assays. LI set up the in-house ELISA assays. CK produced the Cit-C1 and C1 peptides. LP performed the *HLA-DRB1* and *PTPN22* genotyping. RH had main responsibility for the production of Cit-C1 and C1 peptides. LA supervised the statistical analyses (association and interaction analyses) and, together with LK, had main responsibility for the EIRA case-control study. LK and KL designed the study. All authors contributed to the final paper.

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REFERENCES

1. Stastny P. Mixed lymphocyte cultures in rheumatoid arthritis. *J Clin Invest* 1976;**57**:1148–57.
2. Klareskog L, Forsum U, Scheynius A, *et al*. Evidence in support of a self-perpetuating HLA-DR-dependent delayed-type cell reaction in rheumatoid arthritis. *Proc Natl Acad Sci USA* 1982;**79**:3632–6.
3. Gregersen PK, Silver J, Winchester RJ. The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. *Arthritis Rheum* 1987;**30**:1205–13.
4. Klareskog L, Stolt P, Lundberg K, *et al*. A new model for an etiology of rheumatoid arthritis: smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. *Arthritis Rheum* 2006;**54**:38–46.
5. Huizinga TW, Amos CI, van der Helm-van Mil AH, *et al*. Refining the complex rheumatoid arthritis phenotype based on specificity of the *HLA-DRB1* shared epitope for antibodies to citrullinated proteins. *Arthritis Rheum* 2005;**52**:3433–8.
6. Linn-Rasker SP, van der Helm-van Mil AH, van Gaalen FA, *et al*. Smoking is a risk factor for anti-CCP antibodies only in rheumatoid arthritis patients who carry *HLA-DRB1* shared epitope alleles. *Ann Rheum Dis* 2006;**65**:366–71.
7. Makrygiannakis D, Hermansson M, Ulfgren AK, *et al*. Smoking increases peptidylarginine deiminase 2 enzyme expression in human lungs and increases citrullination in BAL cells. *Ann Rheum Dis* 2008;**67**:1488–92.
8. Klareskog L, Rönnelid J, Lundberg K, *et al*. Immunity to citrullinated proteins in rheumatoid arthritis. *Annu Rev Immunol* 2008;**26**:651–75.
9. Snir O, Widhe M, von Spee C, *et al*. Multiple antibody reactivities to citrullinated antigens in sera from patients with rheumatoid arthritis: association with *HLA-DRB1* alleles. *Ann Rheum Dis* 2009;**68**:736–43.
10. Ioan-Facsinay A, el-Bannoudi H, Scherer HU, *et al*. Anti-cyclic citrullinated peptide antibodies are a collection of anti-citrullinated protein antibodies and contain overlapping and non-overlapping reactivities. *Ann Rheum Dis* 2011;**70**:188–93.
11. Mahdi H, Fisher BA, Källberg H, *et al*. Specific interaction between genotype, smoking and autoimmunity to citrullinated alpha-enolase in the etiology of rheumatoid arthritis. *Nat Genet* 2009;**41**:1319–24.
12. van der Woude D, Alemayehu WG, Verduijn W, *et al*. Gene-environment interaction influences the reactivity of autoantibodies to citrullinated antigens in rheumatoid arthritis. *Nat Genet* 2010;**42**:814–6.
13. Verpoort KN, Cheung K, Ioan-Facsinay A, *et al*. Fine specificity of the anti-citrullinated protein antibody response is influenced by the shared epitope alleles. *Arthritis Rheum* 2007;**56**:3949–52.
14. Van PY, Riha GM, Cho SD, *et al*. Blood volume analysis can distinguish true anemia from hemodilution in critically ill patients. *J Trauma* 2011;**70**:646–51.
15. Willemze A, van der Woude D, Ghiddey W, *et al*. The interaction between HLA shared epitope alleles and smoking and its contribution to autoimmunity against several citrullinated antigens. *Arthritis Rheum* 2011;**63**:1823–32.
16. Stolt P, Bengtsson C, Nordmark B, *et al*. Quantification of the influence of cigarette smoking on rheumatoid arthritis: results from a population based case-control study, using incident cases. *Ann Rheum Dis* 2003;**62**:835–41.
17. Plenge RM, Padyukov L, Remmers EF, *et al*. Replication of putative candidate-gene associations with rheumatoid arthritis in > 4,000 samples from North America and Sweden: association of susceptibility with *PTPN22*, *CTLA4*, and *PADI4*. *Am J Hum Genet* 2005;**77**:1044–60.
18. Arnett FC, Edworthy SM, Bloch DA, *et al*. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum* 1988;**31**:315–24.
19. Snir O, Widhe M, Hermansson M, *et al*. Antibodies to several citrullinated antigens are enriched in the joints of rheumatoid arthritis patients. *Arthritis Rheum* 2010;**62**:44–52.
20. Burkhardt H, Koller T, Engström A, *et al*. Epitope-specific recognition of type II collagen by rheumatoid arthritis antibodies is shared with recognition by antibodies that are arthritogenic in collagen-induced arthritis in the mouse. *Arthritis Rheum* 2002;**46**:2339–48.

21. **Burkhardt H**, Sehnert B, Bockermann R, *et al*. Humoral immune response to citrullinated collagen type II determinants in early rheumatoid arthritis. *Eur J Immunol* 2005;**35**:1643–52.
22. **Rothman KJ**, Greenland S, Walker AM. Concepts of interaction. *Am J Epidemiol* 1980;**112**:467–70.
23. **Hosmer DW**, Lemeshow S. Confidence interval estimation of interaction. *Epidemiology* 1992;**3**:452–6.
24. **Dobritzsch D**, Lindh I, Uysal H, *et al*. Crystal structure of an arthritogenic anticollagen immune complex. *Arthritis Rheum* 2011;**63**:3740–8.
25. **Lundberg K**, Kinloch A, Fisher BA, *et al*. Antibodies to citrullinated alpha-enolase peptide 1 are specific for rheumatoid arthritis and cross-react with bacterial enolase. *Arthritis Rheum* 2008;**58**:3009–19.
26. **Kinloch A**, Lundberg K, Wait R, *et al*. Synovial fluid is a site of citrullination of autoantigens in inflammatory arthritis. *Arthritis Rheum* 2008;**58**:2287–95.
27. **Vossenaar ER**, Després N, Lapointe E, *et al*. Rheumatoid arthritis specific anti-Sa antibodies target citrullinated vimentin. *Arthritis Res Ther* 2004;**6**:R142–50.
28. **Masson-Bessière C**, Sebbag M, Girbal-Neuhauser E, *et al*. The major synovial targets of the rheumatoid arthritis-specific antilaggrin autoantibodies are deaminated forms of the alpha- and beta-chains of fibrin. *J Immunol* 2001;**166**:4177–84.
29. **Takizawa Y**, Suzuki A, Sawada T, *et al*. Citrullinated fibrinogen detected as a soluble citrullinated autoantigen in rheumatoid arthritis synovial fluids. *Ann Rheum Dis* 2006;**65**:1013–20.
30. **Pratesi F**, Tommasi C, Anzilotti C, *et al*. Deaminated Epstein-Barr virus nuclear antigen 1 is a target of anti-citrullinated protein antibodies in rheumatoid arthritis. *Arthritis Rheum* 2006;**54**:733–41.
31. **Scherer HU**, van der Woude D, Willemze A, *et al*. Distinct ACPA fine specificities, formed under the influence of HLA shared epitope alleles, have no effect on radiographic joint damage in rheumatoid arthritis. *Ann Rheum Dis* 2011;**70**:1461–4.
32. **Lundberg K**, Alfredsson L, Kallberg H, *et al*. Reply to “Gene-environment interaction influences the reactivity of autoantibodies to citrullinated antigens in rheumatoid arthritis”. *Nat Genet* 2010;**42**:816.



Genetic and environmental determinants for disease risk in subsets of rheumatoid arthritis defined by the anticitrullinated protein/peptide antibody fine specificity profile

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